

[Note]

Development and Application of Simultaneous Analytical Method for Perfluorinated Compounds in Waste Samples by LC/MS/MS

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LC/MS/MS を用いた廃棄物中における有機フッ素化合物 (PFCs) の

一斉分析法の開発及び適用

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The investigation was carried out to establish the simultaneous and high sensitive analysis for the perfluorinated compounds (PFCs) in waste sample.

In pretreatment of the waste, the extraction efficiency of three techniques; accelerated solvent extractor (ASE), soxhlet extraction and ultrasonic extraction were compared and examined.

Operation blank test, recovery test spiked with mass labeled PFCs (MPFCs) and content examination in waste sample were experimented for 14 kinds PFCs.

As for the pretreatment method, the ultrasonic extraction method which had shown good results both in the operation blank and isotope spiked recovery tests were adopted. The PFCs in waste were extracted with methanol and separated as well as concentrated with nitrogen-gas and analyzed by LC/MS/MS determination. And the waste was used in solid form. PFCs were detected at retention time between 3.75~7.15 min by the technique which was established in this study. In addition, the chromatogram of waste sample shows that PFCs and isomer were detected at almost the same time without interference. Since the existence ratios of PFCs in various wastes are different, by establishing profile of each waste at different sources, this study will contribute to developing countermeasures that would prevent environmental contamination of PFCs at global level.

INTRODUCTION

The fluorochemical compounds such as perfluorooctanoate (PFOA) and perfluorooctane sulfonate (PFOS) are stain and water repellent. It is these superior properties that are widely applied to various fields in the modern society for pursuing comfortable and convenient life styles.

On the other hand, negative properties of PFCs such

as hazardous, persistent and bioaccumulative properties have been concerned¹⁾. Thus, in May 9th, 2009, the 4th Conference of the Parties (COP4) for the Stockholm Convention on Persistent Organic Pollutants adopted perfluorooctane sulfonate and perfluorooctane sulfonyl fluoride (PFOS and PFOSF) in Annex B. In addition, these PFOS and PFOSF were designated as a class I specified chemical substances under the Law Concerning the Examination and

Regulation of Manufacture, etc., of Chemical Substances in Japan, and were subject to global regulation. Since the further production and use of these compounds will be strictly controlled, the widespread of the compounds into the environment would be reduced. However, PFCs products have still been widely stocked and used as well as disposed and discharged into our living environment. It is highly likely that the compounds will continue to present ubiquitously in whole society through the various environmental routes and media.

In recent years, many surveys and risk assessment have been carried out with regard to environmental contamination of PFCs in various media including atmosphere and water^{2) -9)}. However, little information has been available with regard to the waste generated as a result of anthropogenic activities and transported to waste treatment facilities. PFCs are detected in high concentrations from leachate at some final disposal sites^{10) 11)}. Thus, the investigation of PFCs in waste will be an important information source to develop countermeasures for PFCs pollution. In this study, sensitive method by using LC/MS/MS for the determination of PFCs in waste samples was developed and presented.

MATERIALS AND METHODS

1. Target PFCs, Reagents and Reagent preparation

The target substances of the development of our analytical method are 14 compounds. The target PFCs and octanol-water partition coefficient ($\log K_{ow}$) are shown in Table 1. The target PFCs in this study were perfluorocarboxylic acids (PFCAs):10, perfluoro alkyl sulfonates (PFASs): 4 substances and mass-labeled PFCs (MPFCs) were mass-labeled PFCAs (MPFCAs):7, mass-labeled PFASs (MPFASs):2 for surrogate compounds. The former target PFCs used PFAC-MXB as standard substances, and the latter MPFCs used MPFAC-MXA as internal standard substances, these were all commercially available products. The both mixture solutions were manufactured by Wellington Laboratories Inc. The PFOA-¹³C₈ used as internal standard was the product of Cambridge Isotope Laboratories. All compounds were linear chains. Methanol was originally for residual agrochemical analysis use obtained from Wako Junyaku Kogyo (Wako Pure Chemical Industries, Ltd.). No interference of the reagents with the measurement of substances was confirmed. The 100ug/L-standard solution for the calibration curve was prepared with 2000ug/L-PFAC-MXB in methanol.

Table 1 Target compounds (PFCs) and octanol-water partition coefficient ($\log K_{ow}$)

| | compound | abbreviation | carbon number | structural formula | molecular weight | $\log K_{ow}$ |
|-------|-----------------------------|--------------|---------------|---|------------------|---------------|
| PFCAs | Perfluoropentanoic acid | PFPeA | 5 | CF ₃ (CF ₂) ₃ COOH | 264.0 | n.a. |
| | Perfluorohexanoic acid | PFHxA | 6 | CF ₃ (CF ₂) ₄ COOH | 314.1 | 3.26 |
| | Perfluoroheptanoic acid | PFHpA | 7 | CF ₃ (CF ₂) ₅ COOH | 364.1 | 3.82 |
| | Perfluorooctanoic acid | PFOA | 8 | CF ₃ (CF ₂) ₆ COOH | 414.1 | 4.30 |
| | Perfluorononanoic acid | PFNA | 9 | CF ₃ (CF ₂) ₇ COOH | 464.1 | 4.84 |
| | Perfluorodecanoic acid | PFDA | 10 | CF ₃ (CF ₂) ₈ COOH | 514.1 | 5.30 |
| | Perfluoroundecanoic acid | PFUnDA | 11 | CF ₃ (CF ₂) ₉ COOH | 564.1 | 5.76 |
| | Perfluorododecanoic acid | PFDoDA | 12 | CF ₃ (CF ₂) ₁₀ COOH | 614.1 | 6.41 |
| | Perfluorotridecanoic acid | PFTTrDA | 13 | CF ₃ (CF ₂) ₁₁ COOH | 664.1 | n.a. |
| | Perfluorotetradecanoic acid | PFTeDA | 14 | CF ₃ (CF ₂) ₁₂ COOH | 714.1 | n.a. |
| PFASs | Perfluorobutane sulfonate | PFBS | 4 | CF ₃ (CF ₂) ₃ SO ₃ H | 300.1 | n.a. |
| | Perfluorohexane sulfonate | PFHxS | 6 | CF ₃ (CF ₂) ₅ SO ₃ H | 400.1 | n.a. |
| | Perfluorooctane sulfonate | PFOS | 8 | CF ₃ (CF ₂) ₇ SO ₃ H | 500.1 | 5.25 |
| | Perfluorodecane sulfonate | PFDS | 10 | CF ₃ (CF ₂) ₉ SO ₃ H | 600.1 | n.a. |

*n.a.: not available

Table 2 Characterization of waste samples

| Sample No. | 1 | 2 | 3 | 4 | 5 | 6 |
|-------------------------|------------------|--------|-----------------------------|------------------------|--------|--------|
| water content ratio (%) | 37.0 | 11.1 | 34.4 | 0.9 | 52.3 | 57.7 |
| pH | 6.8 | 8.0 | 7.6 | 7.9 | 7.4 | 6.8 |
| appearance | dewatered sludge | sludge | automobile shredder residue | shredded solid residue | sludge | sludge |
| color | black | white | black | gray | brown | white |

2. Waste samples

The details of the characteristics of the waste samples such as water content, pH, and appearance are shown in Table 2. The waste samples are dewatered sludge, automobile shredded residues (ASR), incombustible general solid wastes generated from effluent treatment facilities and incinerating process.

3. Equipment and instrumentation, LC/MS/MS analytical condition

The solid phase extract cartridge adopted Precep-C agri (short) (Wako Pure Chemical Industries, Ltd.). The target measurement substances were passed through the cartridge and adsorbed to the cartridge using a Sep-Pak Concentrator System Controller Plus (Waters) by 10 mL/min. The extraction solution was concentrated in TurboVap LV (Zymark) with nitrogen. Analytical conditions of LC/MS/MS are shown in Table 3. As for the ultrasonic generator, ULTRASONIC AUTOMATIC WASHER (AS ONE Corporation) was used. A model 05PR-22 (Hitachi) was used as the centrifugal separator. DIONEX ASE-200 (DIONEX) was selected for the Accelerated Solvent Extractor (ASE). The pulverized wastes were prepared by vibrating sample mill (TI-100, HEIKO Co.), and sieved with 0.5mm-mesh after drying overnight at 50 degrees Celsius.

4. Development of PFCs analysis technique in waste and Examination of the preprocessing method

The following three extraction methods were compared and discussed for sample pretreatment: ASE, soxhlet extraction in Methanol for 12hours and ultrasonic extraction with Methanol. In each

procedure, the operation blank test without interference from the waste and recovery test using real waste samples spiked with 1ng MPFCs were carried out at the same time. In preparing samples, 1ng of MPFCs was spiked to the waste sample as surrogate.

The sample solutions extracted by ASE or soxhlet extraction were concentrated to about 1ml with nitrogen injection. The PFCs adsorbed on the cartridge were then eluted with 2mL methanol. The eluent was concentrated to about 1mL using the TurboVap LV. The pretreated solution extracted by ultrasonic extraction was measured up to 10ml and TurboVap LV. The pretreated solution extracted by ultrasonic extraction was measured up to 10ml and 1mL aliquot of the 10ml extract was taken for determination. Then 1ng of PFOA-¹³C₈ standard solution was spiked as internal standard to the obtained extract, as described above. And 5µL was injected into the LC/MS/MS for determination.

For the measurement of minimum detection limit (MDL), sample 6 which contains little or no PFCs was taken. The waste was spiked with standard mixture methanol solution and adjusted to the final concentration of 2ng/g. The recovery test was repeated for 7 times and the results were used for calculating MDL by using the following equation.

$$MDL = 2 \times s \times t(n-1,0.05)$$

s : standard deviation

t(n-1,0.05) : t value of n-1 degree of freedom, level of significance 5%

Table 3 Analytical condition for PFCs with LC/MS/MS

| LC condition | | | | | | |
|----------------------|---------------------------------------|--------------|-----------------------------|------------|--------|----------------|
| Instrument | ACQUITY UPLC | | (waters) | | | |
| Column | UPLC BEH C18 | | 2.1×50mm | | | |
| Retention gap Column | UPLC BEH C18 | | 2.1×100mm | | | |
| Mobile Phase | A : 10mM Ammonium | | Acetate aq B : Acetonitrile | | | |
| Gradient | 0.0 | 8.0 min | B: 1 95% | | | |
| | 8.0 | 8.1 min | B: 95 1% | | | |
| Flow rate | 0.3 mL / min | | | | | |
| Column temp. | 50 | | | | | |
| Injection volume | 5µL | | | | | |
| MSMS condition | | | | | | |
| Instrument | ACQUITY TQD | | (waters) | | | |
| Ionization Mode | ESI (-) | | Source temp. | 120 | | |
| Desolvation temp | 300 | | Cappillary voltage | 2kV | | |
| Cone gas flow | 20 L / hr | | Desolvation gas flow | 800 L / hr | | |
| Collision gas flow | 0.1 mL / min | | | | | |
| | | | | | | |
| | Compound name | Precursor io | Product ion | | Cone | Collision |
| | | | Qnt. | Idt. 1 | Idt. 2 | Voltage Energy |
| PFCAs | PFPeA | 263.00 | 219.00 | | | 18 9 |
| | PFHxA | 313.00 | 269.00 | 118.90 | | 18 9 |
| | PFHpA | 363.00 | 318.90 | 169.00 | | 18 11 |
| | PFOA | 413.00 | 368.90 | 169.00 | | 18 11 |
| | PFNA | 463.00 | 418.90 | 169.00 | | 18 13 |
| | PFDA | 513.00 | 468.90 | 219.00 | | 21 13 |
| | PFUdDA | 563.00 | 518.80 | 269.00 | | 18 15 |
| | PFDoDA | 613.00 | 568.90 | 168.90 | | 24 15 |
| | PFTTrDA | 663.00 | 618.80 | 169.00 | | 24 17 |
| | PFTeDA | 713.00 | 668.70 | 169.00 | | 24 15 |
| MPFCAs | PFBA - ¹³ C ₄ | 217.00 | 172.00 | | | 15 13 |
| | PFHxA - ¹³ C ₂ | 315.00 | 270.00 | | | 15 11 |
| | PFOA - ¹³ C ₄ | 417.00 | 371.90 | | | 27 11 |
| | PFNA - ¹³ C ₅ | 467.00 | 422.90 | | | 24 15 |
| | PFDA - ¹³ C ₂ | 515.00 | 469.80 | 219.00 | | 21 14 |
| | PFUdDA - ¹³ C ₂ | 565.00 | 519.80 | 269.60 | | 21 17 |
| | PFDoDA - ¹³ C ₂ | 615.00 | 569.90 | 269.50 | | 24 14 |
| PFASs | PFBS | 299.00 | 79.90 | 98.90 | | 51 37 |
| | PFHxS | 399.00 | 79.90 | 98.90 | 130.00 | 57 46 |
| | PFOS | 499.00 | 79.90 | 98.90 | 130.00 | 69 55 |
| | PFDS | 599.00 | 79.90 | 98.90 | 79.90 | 80 67 |
| MPFASs | PFHxS - ¹⁸ O ₂ | 403.00 | 83.90 | 103.00 | | 55 47 |
| | PFOS - ¹³ C ₄ | 503.00 | 79.90 | 99.00 | | 65 56 |

RESULTS AND DISCUSSION

1. Result of the pretreatment procedure

The results of operation blank by using the 3 methods are shown in Fig.1. High concentrations of PFHxA and PFHpA in PFCAs that exceeded 7ng/g-wet were detected by the ASE and soxhlet extraction. The result confirmed cross contamination in the operation process. The concentrations of the other PFCs and those of the PFCs extracted by ultrasonic extraction method were under 1ng/g-wet. The contamination level of PFCs was not as significant as to affect the analysis seriously. The result of the recovery test of the samples from 1 to 4 spiked with 1ng MPFCs is shown in Fig.2. The 3 color bar charts show the average recovery rate. The recovery rate of the MPFHxA and MPFDoA by the ASE, soxhlet extraction, as well as MPFHxS by the ASE were under 60%, while the rest of the recovery rates show no significant differences among the extraction methods. Despite the use of wastes in various forms and properties as target samples, the recovery rates of the samples from 1 to 4 were not significantly affected by the extraction methods used in the study. It is, therefore, concluded that extraction can be performed by either technique without a problem.

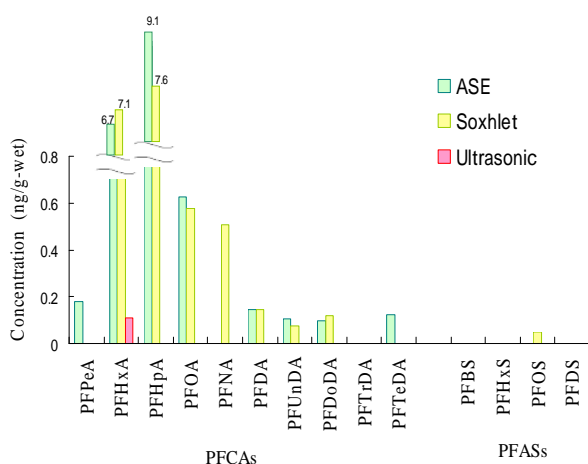


Fig.1 PFCs concentration of operational blank

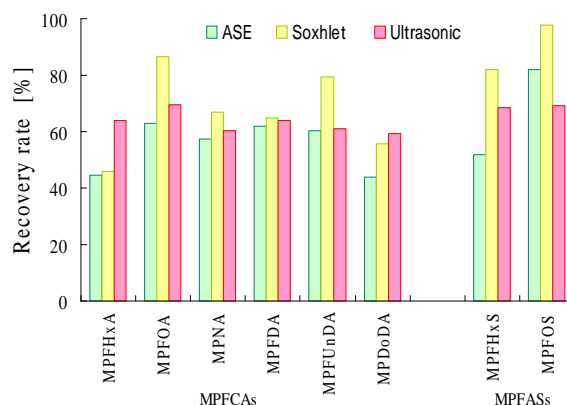


Fig.2 Recovery rate with MPFCs

2. Content examination in the waste sample

The results of content in waste sample1-4 with every method to compare extraction efficiency show in Fig.3.

The PFCs content were indicated to be deducted with the content obtained by the operation blank. In ultrasonic extraction method, each sample were analyzed repeatedly 3 times and showed the range of concentration (black bar line), the average (red bar charts). PFCs were detected from all waste samples.

As an example of PFCs analysis, the PFCAs chromatograms of the standard solution (20ng/L) and the sample1 are shown in Fig.4, and the PFASs chromatograms are shown in Fig.5, respectively. In the chromatogram of standard solution (left), PFCAs was detected between 3.75~7.15 min. The chromatograms of waste sample (right) show that PFCAs and isomer were detected almost at the same time without interferences. And the minimum detection limit values of PFCs obtained by ultrasonic extraction method were 1.4 - 3.9ng/g.

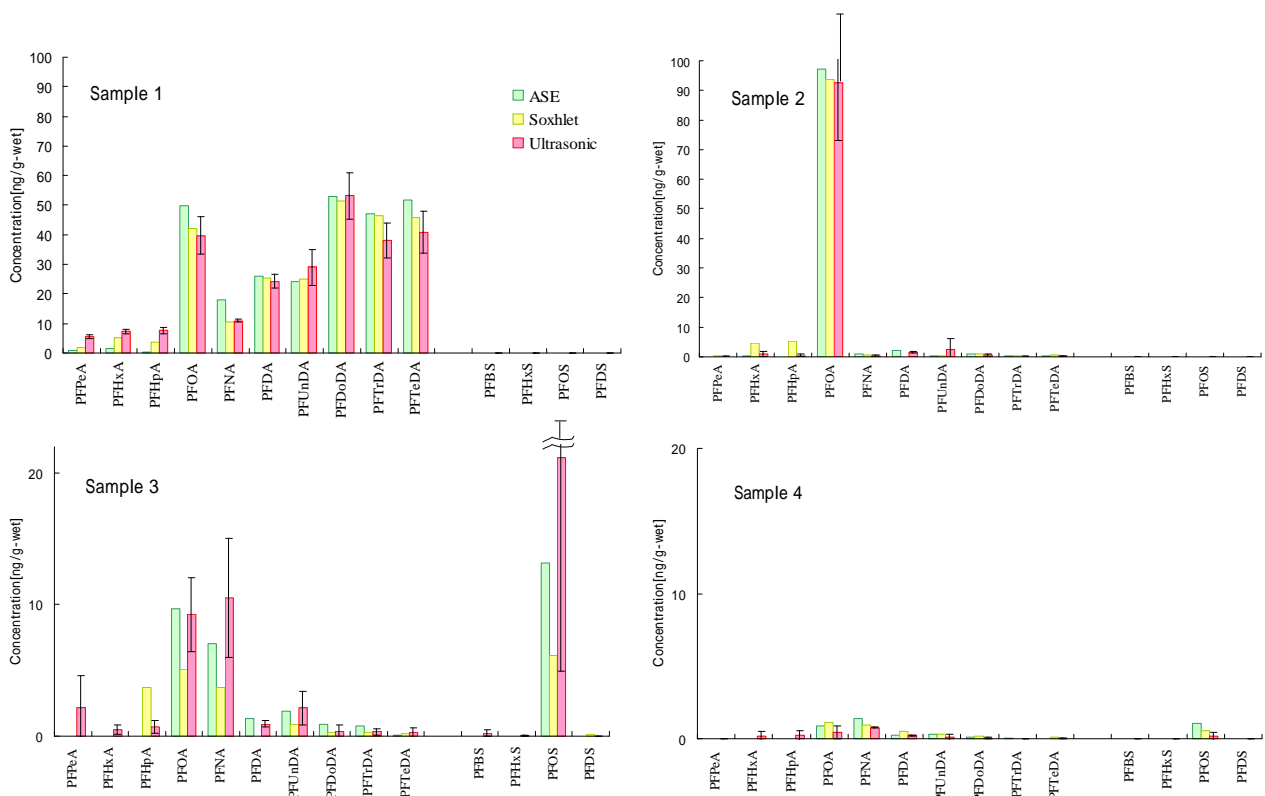


Fig.3 PFCs concentration in waste sample comparing extraction between different 3 methods

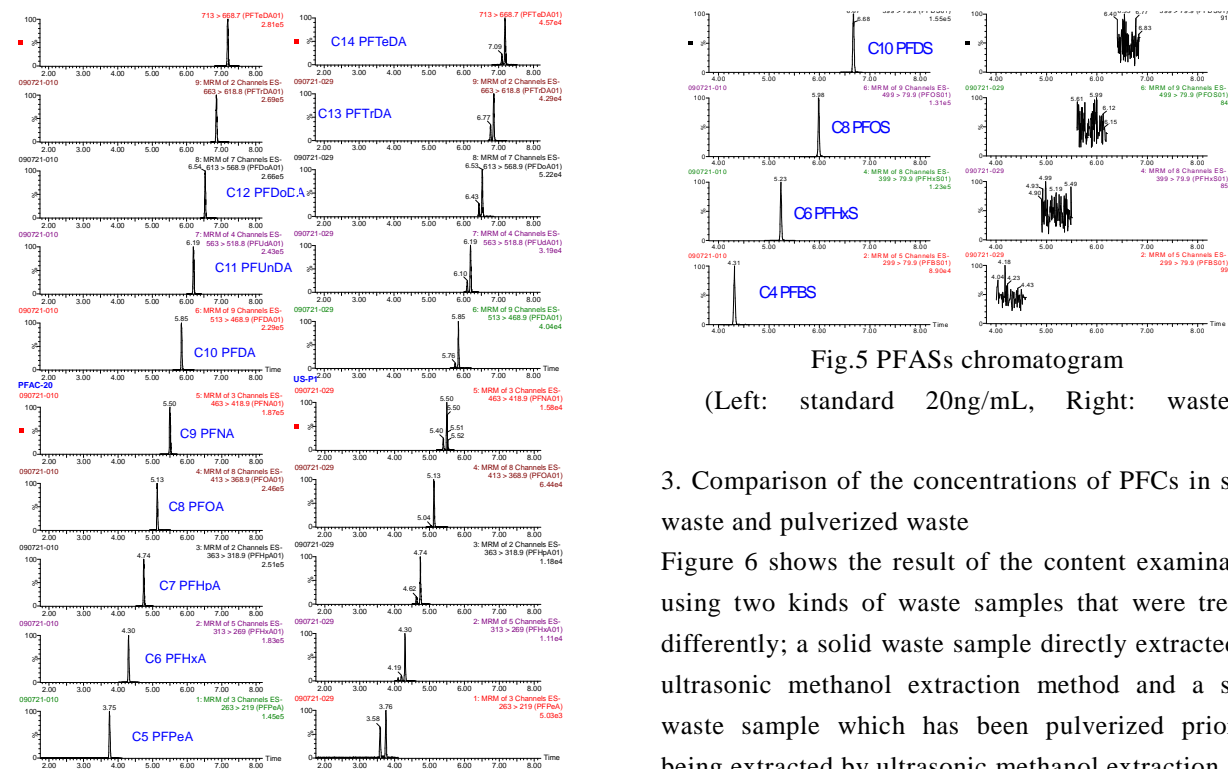


Fig.4 PFCAs chromatogram (Left: standard 20ng/mL, Right: waste sample 1)

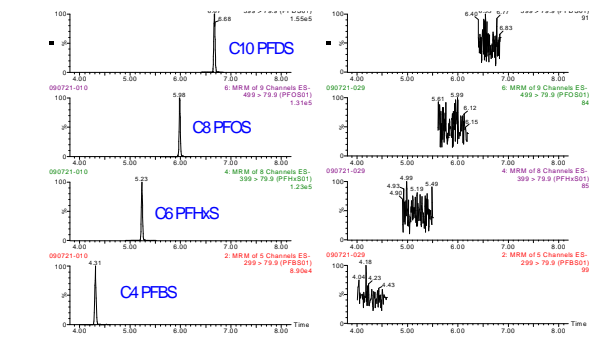


Fig.5 PFASs chromatogram (Left: standard 20ng/mL, Right: waste)

3. Comparison of the concentrations of PFCs in solid waste and pulverized waste

Figure 6 shows the result of the content examination using two kinds of waste samples that were treated differently; a solid waste sample directly extracted by ultrasonic methanol extraction method and a solid waste sample which has been pulverized prior to being extracted by ultrasonic methanol extraction method, as an example, sample1 and sample3. The concentrations of PFCs in the solid sample and the pulverized sample were nearly the same for waste sample1 (dewatered sludge). On the other hand, the waste sample3 (ASR) shows higher concentrations of PFCs in the solid state than the pulverized one.

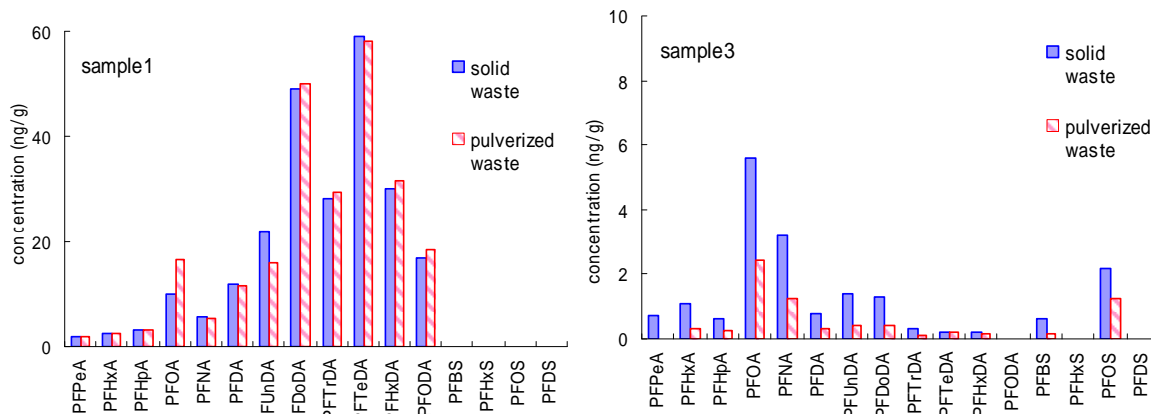


Fig.6 Comparison solid waste with pulverized waste

CONCLUSIONS

The analysis method for PFCs in waste samples was established.

Ultrasonic extraction method was employed, which gave good results as a pretreatment method, for both operation blank and isotope recovery tests. These analytical procedures are composed by methanol extraction, centrifugation, concentration with nitrogen-gas and determination by LC/MS/MS. And the waste was used in solid form.

PFCs were detected at retention time between 3.75~7.15 min without interferences.

MDL was 1.4 - 3.9ng/g. It was confirmed that this method was applicable for PFCs analysis in waste samples. As the result, the difference existence ratios of PFCs in various wastes was shown, establishing profile of each waste at different sources became possible.

The result indicates that the existence ratios of PFCs in various wastes are different, which leads to the possible establishment of a profile of each waste generated from different sources.

REFERENCES

1)Kannan K., Corsolini S., Falandysz J., Fillmann G., Kumar KS., Loganathan BG, Mohd MA, Olivero J, Van Wouwe N., Ho Yang J.,Aldous K.M. : Perfluorooctanesulfonate and related fluorochemicals in human blood from several countries, *Environ. Sci. Technol.*, **38**, 4489-4495(2004)

2)Murakami M., Takizawa S. : Current Status and Future Prospects of Pollution in Water Environments by perfluorinated surfactants, *Journal of Japan Society on Water Environment*, **33(8)**, 103-114 (2010)

3)Harada K., Saito N.,Inoue K., Yoshinaga T., Watanabe T., Sasaki S., Kamiyama S., Koizumi A. : The Influence of Time, Sex and Geographic Factors on Levels of perfluorooctane sulfonate and perfluorooctanoate in human serum over the Last 25 years, *J. of Occupational Health*, **46**, 141-147(2004)

4)Lien N.P.H., Fujii S., Tanaka S. : PFOS and PFOA contamination in Surface Water and Drinking Water in Kinki Area, The 41st Annual Conference of Japan Society on Water Environmental, 497(2007)

5)Sasaki K., Yaegashi K., Saito N. : Perfluorooctane sulfonate(PFOS) and perfluorooctanoate(PFOA), Environmental Monitoring Report 2004: 37-59(2004)

6)Yoshida M., Kobuke Y., Nakano T. : Contamination of PFOA and PFOS in the rivers of Hyogo pref. Japan, *Organohalogen Compounds*,**69**, 2881-2884(2007)

7)Matsumura C., Takemine S., Yoshida M., Nakano T. : Contamination of perfluorinated compounds in water environment of Hyogo prefecture, *18th Symposium on Environmental Chemistry Programs and Abstracts*, 592-593 (2009)

8)Matsumura C., Takemine S., Yoshida M., Suzuki M., Yamamoto A., Tsurukawa M., Nakano T. : Contamination of perfluorinated compounds in the river water of Hyogo pref. Japan, *Organohalogen Compounds*,**71**, 2315-2319(2009)

9)Yoshida M., Matsumura C., Takemine S., Okuno T., Nakano T., Yamamoto A., Tokai A., Morioka T. : Risk assessment of PFOA, PFOS and determination of perfluorinated compounds in blood, *Organohalogen Compounds*, **71**, 2605-2608(2009)

10)Yagi M., Yamaji A., Shibutani I. : Tendency of fluorinated organic compounds in the process of leachate disposal of final landfill site, 18th *Symposium on Environmental Chemistry Programs and Abstracts*, 558-559 (2009)

11)Kurihara M., Yoshizawa T., Oishi O., Fujimura Y., Kojima H. : Study of PFCs in water treatment of disposal site, 19th *Symposium on Environmental Chemistry Programs and Abstracts*, 508-509(2010)

要 約

本研究は、廃棄物中において、高感度な PFCs 一斉分析手法を確立するために実施した。廃棄物の前処理法には、ASE、ソックスレー、超音波抽出法の3手法を比較検討した。14種のPFCsについて、操作ブランク試験、同位体を用いた添加回収試験、固形及び粉碎廃棄物を用いた含有量試験などを実施した。

その結果、前処理法としては、操作ブランク、同位体回収試験で良好な結果であった超音波抽出法を採用した。即ち、廃棄物中PFCsは、メタノール抽出、遠心分離、窒素ガスでの濃縮、LC/MS/MSで分析した。また、廃棄物の形状は、固形状のまま用いることとした。本研究で確立した手法で廃棄物の含有量を測定したところ、妨害もなく、保持時間3.75分~7.15分の間を検出できることを確認した。様々な廃棄物中のPFCsの存在割合は異なり、排出源の異なる廃棄物ごとのプロファイルを確立することで汚染防止対策に寄与できると期待される。